## HCR In situ hybridization protocol, Kit Version 3

Adapted From: Choi, H.M.T., M. Schwarzkopf, M.E. Fornace, A. Acharya, G. Artavanis, J. Stegmaier, A. Cunha, and N.A. Pierce. 2018. Third-generation in situ hybridization chain reaction: multiplexed, quantitative, sensitive, versatile, robust. BioRxiv, doi: https://doi.org/10.1101/285213. Available at: https://www.biorxiv.org/content/early/2018/03/19/285213

## Fixation and Dehydration of Specimens

- 1. Dechorinate embryos with fine tweezers or a pair of needles.
- 2. Transfer embryos to an eppendorf tube and fix them with 1 mL of 4 %paraformaldehyde and keep them at 4 °C for at least 24 hours.
- 3. Wash embryos 3 times for 5 minutes with 1mL of 1X PBS (**DEPC treated/RNase free**) to stop fixation. Fixed embryos can be stored at 4°C at this stage.
- 4. Dehydrate embryos in a series of methanol (MeOH) washes with 1 mL each (100% MeOH 4 times for 10 min and 100% MeOH once for 50 min
- 5. Store embryos at -20 °C.

## Rehydrate specimens with a series of 1 mL MeOH/PBST washes

- 1. Set-up 6 washes
  - a. 3:1 MeOH: 1X PBST 5 min
  - b. 1:1 MeOH: 1X PBST 5 min
  - c. 1:3 MeOH: 1X PBST 5 min
  - d. 100% 1X PBST 5 min three times

#### Proteinase K

- 1. Treat embryos with 5  $\mu$ L of PK in 100  $\mu$ L of 1X PBST for 10 min at room temperature.
- 2. Stop PK by replacing the solution with 250  $\mu$ L of 4% PFA in 100  $\mu$ L of PBST for 20 min at room temperature.
- 3. Wash embryos twice in 250  $\mu$ L of 1X PBST for 5 min at room temperature.

(While the washes are going heat up the water bath to 37  $^{\circ}$ C and warm up 250  $\mu$ L of probe hybridization buffer)

## **Probe Hybridization**

- 1. Pre-hybridize the embryos in 250 μL of 30% of probe hybridization buffer for 3 hours at 37°C.
- 2. Prepare probe solution by adding 2 pmol of each probe solution (odd and even:  $0.5~\mu L$  of 2  $\mu M$  stock per probe mixture to 250  $\mu L$  of 30% probe hybridization buffer at 37°C).
- 3. After 3 hours the embryos are finished pre-hybridizing.
- 4. Remove the pre-hybridization solution and add the probe solution.

Written by Alexander Jagla - June 2016; Modified by Sara Alharbi & Windsor Aguirre May 2018.

- 5. Incubate embryos in probe solution for at least (12-16) hours at 37°C in the dark.
- 6. Remove the excess probes by washing with 250  $\mu$ L of the following solutions at 37°C (**Pre-heat wash solutions to 37°C before use**)
  - a. Wash embryos 4 times for 15 minutes with 250  $\mu$ L of 30% probe wash buffer at 37 °C
- 7. Wash embryos 3 times for 5 minutes with 5X SSCT at room temperature.

# Amplification (When working with hairpin stocks remember to change tips between stock tubes! Do not contaminate the amplifiers! Keep embryos and amplifiers in the dark.)

- 1. Pre-amplify the embryos in 250  $\mu$ L of amplification buffer for **30 min** at room temperature.
- 2. Prepare 30 pmol of each fluorescently labeled hairpin by snap cooling 5  $\mu$ L of 3  $\mu$ M stock in hairpin storage buffer (heat at 95°C for 90 seconds and cool to room temperature in a dark for 30 min).
- 3. Prepare hairpin solution by adding all snap-cooled hairpins to 250 µL of amplification buffer at room temperature.
- 4. Replace pre-amplification buffer with the hairpin and amplification buffer solution.
- 5. Incubate embryos in the amplifiers for at least (12-16) hours at room temperature in the dark.
- 6. Remove excess probes by washing at room temperature with 250  $\mu$ L of the following solutions:
  - a. Wash twice in 5X SSCT for 5 min.
  - b. Wash twice in 5X SSCT for 30 min.
  - c. Wash one time in 5X SSCT for 5 min.

### **Mounting Embryos**

- 1. Mount embryos in depression slides (or use slides with a glass ring, nylon washer, or well made out of scotch tape).
- 2. Apply a drop of Aquamount or similar mounting solution.
- 3. Using a transfer pipet, pipet an embryo into the mounting solution.
- 4. Orient the embryo in the mounting solution using a nylon loop (e.g. fishing line) or similar.
- 5. Attach a glass coverslip.

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# **Reagents**

- -4% Paraformaldehyde (store at 4°C)
- -Methanol
- -Proteinase K
- -1X PBS
- -1X PBST
- -5X SSCT
- -5X SSC
- -Probe Hybridization Buffer (Molecular Instruments)
- -Probe Wash Buffer (Molecular Instruments)
- -Amplification Buffer (Molecular Instruments)
- -DNA Probes (Molecular Instruments)
- -Amplifiers (Molecular Instruments)
- -Aquamount (Fisher Catnum: 14-390-5)